

The Ameliorative Effects of Ethyl Pyruvate and Dimethyl Sulfoxide on Ischemic Tissue Injury in Experimental Carbon Monoxide Intoxication

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Abstract

Aim: The purpose of this study was to investigate the protective effects of ethyl pyruvate (EP) and dimethyl sulfoxide (DMSO) on ischemic tissue in brain and cardiac and hepatic tissues in experimentally induced carbon monoxide (CO) intoxication.

Material and Methods: Thirty-five mature female Sprague Dawley rats were randomized into five groups of seven animals each. Group 1 received no CO or treatment. Rats in groups 2, 3, 4 and 5 were made to inhale a high-concentration 5000 ppm CO gas mixture for 60 min at 4 L/min. CO levels were then measured from 1 mL tail vein blood. Group 2 received no therapeutic agent, group 3 received 6 mg/kg intraperitoneal (ip) DMSO, group 4 received 50 mg/kg ip EP, group 5 received 50 mg/kg EP and 6 mg/kg DMSO ip. All rats were sacrificed by decapitation. Brain, cardiac and hepatic tissues were removed and histopathological scores were compared.

Results: Comparison of group 2 with group 3 and group 2 with group 5 revealed that DMSO alone and EP + DMSO exhibited a reducing effect on degree of cerebral neuronal alteration, degenerative neuron rates, and total cardiac injury score ($p=0.005$, $p=0.002$, $p=0.004$ and $p=0.002$, $p=0.001$, $p=0.004$ respectively). There was no histopathological difference between group 2 and group 4 suggesting EP alone had no therapeutic effect on histopathological injury.

Conclusion: Based on our study findings, EP administered alone exhibited no protective effects on the organ injuries investigated, while DMSO exhibited reducing effects on degree of neuron alteration, rates of degenerative neurons, and total cardiac injury scores. An ameliorating effect on cardiac and hepatic injury was more prominent with combined treatment.

Keywords: Carbon monoxide, dimethyl sulfoxide, ethyl pyruvate, experimental, intoxication

Introduction

Carbon monoxide (CO) is a colorless, odorless, tasteless, non-irritating gas which normally presents at a level of 0.001% in the atmosphere. It emerges when carbon-based compounds used as fuels for cooking and heating are incompletely consumed, or during fires (1). CO is one of the most important global agents in injury-

and intoxication-related mortality (2). Children and the elderly are more rapidly affected. Other factors increasing disposition are exercise, stress, and anemia. High atmospheric concentrations and long-term exposure are other important factors. Due to their high oxygen requirements, brain, heart, and liver are the most susceptible organs to the hypoxic effects of CO exposure (3). Studies have determined that the cardiac effect is associated with



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Cite this article as: Paslı Sinan, İmamoğlu Melih, Çiçek Mustafa, Yadigaroglu Metin, Şahin Aynur, Karaca Yunus, Yuluğ Esin, Tatlı Özgür. The Ameliorative Effects of Ethyl Pyruvate and Dimethyl Sulfoxide on Ischemic Tissue Injury in Experimental Carbon Monoxide Intoxication. Eurasian J Emerg Med. 2020;19(1):30-9.

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Eurasian Journal of Emergency Medicine published by Galenos Publishing House.

Received: 30.08.2019

Accepted: 11.12.2019

formation of carboxyhemoglobin (COHb) which causes hypoxia, and that CO-related oxidative stress leads to neurological injury (4). The synthesis of reactive oxygen products increases secondary to tissue hypoxia in CO intoxication. The numbers of studies of the place and importance of antioxidant therapies as an alternative to oxygen therapy are therefore increasing rapidly. Several studies agree that antioxidant therapy can be a novel therapeutic option in the treatment of CO intoxication (5). Dimethyl sulfoxide (DMSO) is a strong penetration enhancer, cell fusogen and cryoprotectant. It is a widely used molecule in cell biology (6). It is mainly intended for use in many gastrointestinal diseases, thanks to its anti-inflammatory and reactive oxygen species cleaning activities (7). DMSO crosses the blood-brain barrier and has been effective in treating traumatic brain edema (8,9). It has also been used in the treatment of musculoskeletal disorders (10), rheumatologic diseases (11), dermatological diseases and as a topical analgesic (12). In addition, it has been recommended for the treatment of Alzheimer's disease (13). Ethyl pyruvate (EP) is a stable and simple lipophilic ester originating from the endogenous metabolite pyruvate. It has been shown to provide protection against inflammation and reduce organ dysfunction in some models of clinical disease, such as burns, sepsis, and acute pancreatitis (14,15). Shen et al. (16) has shown that ethyl pyruvate improves hypoxic-ischemic brain injury through the anti-inflammatory and anti-cell death mechanism. The effect of ethyl pyruvate on ischemia reperfusion injury has also been demonstrated by scientists. Hu et al. (17) have shown that ethyl pyruvate reduces myocardial ischemia reperfusion injury by inhibiting HMGB1 protein expression in rats. In addition, Tsung et al. (18) has shown that ethyl pyruvate protects against hepatic ischemia / reperfusion (I/R) injury by reducing hepatic necrosis and apoptosis. The purpose of this study was to investigate the protective effects of ethyl pyruvate and DMSO at the histopathological level in experimentally-induced CO intoxication in rats.

Materials and Methods

Study Design

This randomized, controlled, non-blinded, interventional animal study was performed following receipt of approval from Karadeniz Technical University Animal Care and Ethics Committee (approval no: 2016/23). A 5000 ppm experimental CO intoxication protocol was applied when establishing the experimental model (19). Intraperitoneal (ip) administration of 6 mg/kg DMSO and 50 mg/kg EP was planned in the light of previous studies.

Animal Subjects

Thirty-five mature female Sprague Dawley rats (10-week old, weighing 250 ± 25 g) were used in this study. Until the day of the

study, rats were housed in steel cages at 22 °C room temperature with access to water and standard rat chow. The CO gas to be used in the experimental protocol was ordered from HABAŞ Sınai ve Tarım Gazlar Endüstrisi A.Ş. (İzmit, Turkey) such as to contain 5000 pp CO concentrations. A special mechanism consisting of a two-chamber aquarium tank 100x40x50 cm in size, with holes capable of receiving gas from one end and permitting gas to escape from the other was employed for the experiment.

Study Protocol

The 35 rats were randomly assigned into five groups of seven animals each.

Group 1 (Sham group, n=7): After being placed into the glass container, these rats inhaled room air for 60 min, after which CO levels were measured from 1 mL blood gas from the tail veins.

Group 2 (Control group, n=7): After being placed into the glass container, these rats inhaled high-concentration 5000 ppm CO gas mixture at 4 L/min for 60 min, after which CO levels were measured in 1 mL blood gas from the tail veins.

Group 3 (DMSO group, n=7): After being placed into the experimental glass container, these rats inhaled high-concentration 5000 ppm CO gas mixture at 4 L/min for 60 min, after which CO levels were measured in 1 mL blood gas from the tail veins. Immediately after removal from the glass jar, 6 mg/kg DMSO was administered ip.

Group 4 (EP group, n=7): After being placed into the experimental glass container, these rats inhaled high-concentration 5000 ppm CO gas mixture at 4 L/min for 60 min, after which CO levels were measured in 1 mL blood gas from the tail veins. Immediately after removal from the glass jar, 50 mg/kg EP was administered ip.

Group 5 (DMSO plus EP group, n=7): After being placed into the experimental glass container, these rats inhaled high-concentration 5000 ppm CO gas mixture at 4 L/min for 60 min, after which CO levels were measured in 1 mL blood gas from the tail veins. Immediately after removal from the glass jar, 50 mg/kg EP and 6 mg/kg DMSO were administered ip.

Histopathological Examination

All rats were sacrificed by decapitation 24 h after removal from the glass tank. Brain, cardiac, and hepatic tissues were removed. Each tissue was divided into two parts down the midline, ensuring that each part contained all layers. These tissue specimens were fixed in 10% neutral formalin for 48 h. Specimens were dehydrated by being passed through increasingly greater alcohol concentrations, made transparent in xylene, and embedded in paraffin blocks for histopathological examination. Sections 5 µm in thickness were taken from the paraffin block using a

fully automatic microtome (Leica RM 2255, Tokyo, Japan) and stained with hematoxylin-eosin (H&E) for detailed evaluation. Cerebral tissue specimens were also stained with cresyl violet for more detailed examination. Preparates were evaluated by an experienced histologist (E.Y.) blinded to the study groups using a light microscope (Olympus BX-51; Olympus Optical Co., Tokyo, Japan). Preparates were scored semi-quantitatively for all organs (0=none, 1=mild, 2=moderate, 3=severe). Tissues' histopathological damage scores are shown in Table 1 (20,21,22,23). Neuronal alterations in cerebral tissue were scored histologically (20).

Total cardiac and hepatic injury scores were obtained by adding the histopathological parameter scores for cardiac and hepatic

tissues, and statistical comparisons were performed on the basis of these scores.

COHb Measurement

COHb levels were determined using an automatic blood gas measurement device (Rapidlab 1265, Bayer Health Care LLC, Pittsburgh, PA, USA).

Statistical Analysis

Statistical analyses were performed on SPSS 23.0 (IBM SPSS, Armonk, NY, USA) software. Results were expressed as median and 25-75% quartiles. Kruskal-Wallis analysis of variance was used for group comparisons (Mann-Whitney U test with post hoc Bonferroni correction). P values <0.05 were considered statistically significant.

Table 1. Histopathological classification of tissue damage in brain, cardiac, and hepatic tissues
Brain
Neuronal changes (20)
1- Group with mildly shrunken neurons with / without cytoplasmic vacuolization,
2-Moderately shrunken neurons (eosinophilic cytoplasm) and increased nuclear basophilia or vacuoles with cytoplasm and vesicular nucleus,
3-Severely shrunken neurons (eosinophilic cytoplasm), piknotic nucleus
Cells with shrinkage in the cell body, loss of Nissl substance and eosinophilia, and cells with a shrunken nucleus were regarded as degenerative (21)
Cardiac (22)
Myocardial cell degeneration
0-No degeneration in myocytes,
1-Some degenerated myocytes,
2-Approximately 50% myocyte degeneration,
3-More than 50% myocyte degeneration
(Myocytes with multifocal vacular degeneration and piknotic nuclei were regarded as degenerative.)
Edema, vascular congestion, increased spaces between muscle fibers
0-None
1-Mild
2-Moderate
3-Severe
Hepatic
Hepatic injury (23)
0-None or minimal
1-Mild injury with cytoplasmic vacuolization and focal nuclear piknosis,
2-Diffuse nuclear piknosis of variable severity, and loss of cell margins,
3-Severe necrosis with neutrophil infiltration and hemorrhage
Hepatocyte degeneration, vascular congestion, and sinusoidal dilatation (23)
0-None
1-Mild
2-Moderate
3-Severe

Results

Mean COHb concentrations for all groups are shown in Table 2. Although all groups were exposed to CO for the same time and at the same concentrations, CO levels differed significantly among the groups. Analysis revealed no significant differences between group 2, 3, and 4 CO levels among the CO-exposed groups. However, differences were determined between group 5 CO levels and those of groups 2 and 4 ($p=0.033$ and 0.019 , respectively).

Histopathological Evaluation

The study groups' histopathological results and median injury values are shown in Table 2. Kruskal-Wallis analysis performed to

compare the five groups revealed significant differences between them in terms of injury parameters ($p=0.000$). Bonferroni correction was applied to determine which groups representing the source of the variation (Table 3).

Brain tissue evaluation investigated shrinkage in the cell body, loss of Nissl substance and eosinophilia in cytoplasm, and degenerative neurons with a dark, shrunken nucleus. Diffuse pyramidal neurons with a normal architecture were observed in the cerebral cortex in group 1 (Figure 1). Diffuse degenerative neurons were observed in all layers of the cortex in group 2 (Figure 2). Diffuse pyramidal neurons with normal architecture were observed in addition to degenerative neurons in group 3 (Figure 3). Occasional pyramidal neurons with normal architecture in addition to diffuse degenerative neurons in all layers were observed in group 4 (Figure 4). Diffuse pyramidal neurons with normal morphology in addition to degenerative neurons were observed in group 5 (Figure 5). Cardiac tissue evaluation revealed a normal myocardial cell structure in group 1 (Figure 6). Degeneration in cardiac muscle cells and vascular congestion and edema between muscle fibers were observed in group 2 (Figure 7). Cardiac muscle cells in group 3 exhibited a close to normal morphology, and occasional vascular

COHb	Group 1	Group 2	Group 3	Group 4	Group 5
Median	0.350	36.4	29.05	35.850	18.600
Minimum	0.3	25.2	22.7	24	11
Maximum	4.4	47.8	39.9	52.6	33.8
25-75%	0.3-3.4	29.9-42.8	23.3-38.1	28.9-51.9	14.6-28.3
CO: Carbon monoxide, COHb: Carboxyhemoglobin					

Histopathological Parameter	Group 1 (Sham) Median %95 CI	Group 2 (CO) Median 95% CI	Group 3 (DMSO) Median %95 CI	Group 4 (EP) Median 95% CI	Group 5 (EP + DMSO) Median 95% CI	p values
Neuron alterations (Grade)	1.500 0.581-2.419	3.000 2.263-3.166	1.000 0.454-2.046	3.000 2.125-3.209	2.000 1.125-2.209	0.005 $P^{1-2}=0.002$ $P^{1-4}=0.004$ $P^{2-3}=0.005$
Degenerative brain cell rates	11.500 6.834-17.666	65.000 59.246-70.468	25.500 18.069-30.931	55.000 50.519-61.815	27.500 22.074-31.593	0.005 $p^{1-2,3,4,5}=0.002$ $p^{2-3,5}=0.002$ $p^{3-4}=0.002$ $p^{4-5}=0.002$
Total cardiac injury score	2.000 0.163-3.837	6.000 5.077-6.066	4.000 3.454-5.046	5.000 4.249-6.417	2.500 1.399-3.601	0.005 $P^{1-2,3,4}=0.001$ $P^{2-3}=0.004$ $P^{2-5}=0.001$ $P^{4-5}=0.002$
Total hepatic injury score	2.000 0.701-3.299	7.000 5.503-8.211	5.000 3.909-7.091	5.500 4.802-6.865	3.500 2.810-4.524	0.005 $P^{1-2,3,4}=0.001$ $P^{1-5}=0.003$ $P^{2-5}=0.004$ $P^{4-5}=0.002$

CO: Carbon monoxide, EP: Ethyl pyruvate, DMSO: Dimethyl sulfoxide, CI: Confidence interval, DMSO: Dimethyl sulfoxide

DMSO exhibited a greater capacity to reduce neuronal alteration grades, the degenerative neuron rates, and cardiac total injury score in Group 3, compared to Group 2 (CO) ($p=0.005$, $p=0.002$, and $p=0.004$, respectively). Comparison between Group 5 (EP + DMSO) and Group 2 revealed that EP + DMSO reduced the degenerative neuron rate, total cardiac injury score, and total hepatic injury score ($p=0.002$, $p=0.001$, and $p=0.004$, respectively). No significant difference was determined between Group 3 and Group 5 in terms of treatment results

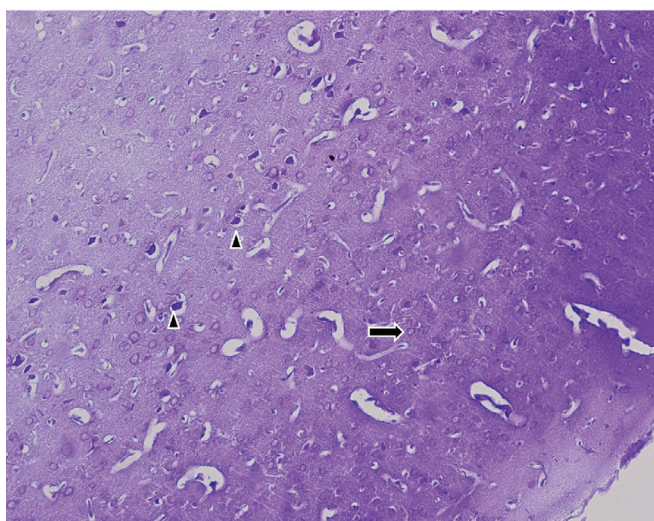


Figure 1. (Group 1) Cerebral cortex microphotographs (cresyl violet X 200). Normal pyramidal neurons (↑), degenerative pyramidal neurons (Δ)

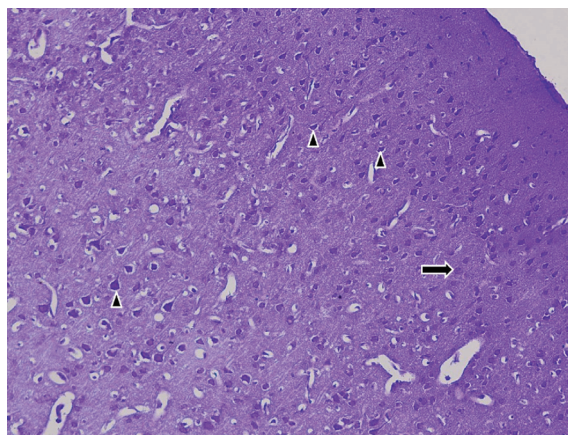


Figure 2. (Group 2) Cerebral cortex microphotographs (cresyl violet X 200). Normal pyramidal neurons (↑), degenerative pyramidal neurons (Δ)

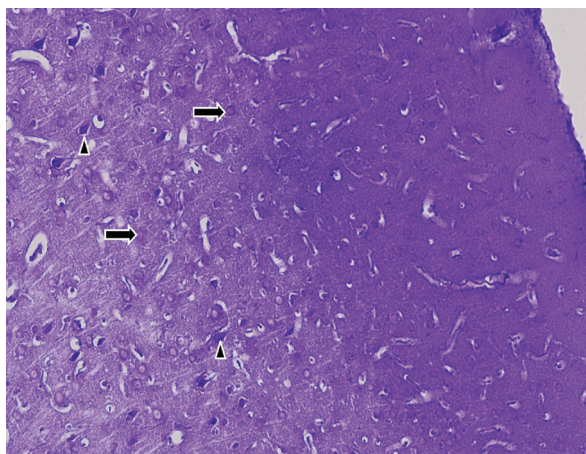


Figure 3. (Group 3) Cerebral cortex microphotographs (cresyl violet X 200). Normal pyramidal neurons (↑), degenerative pyramidal neurons (Δ)

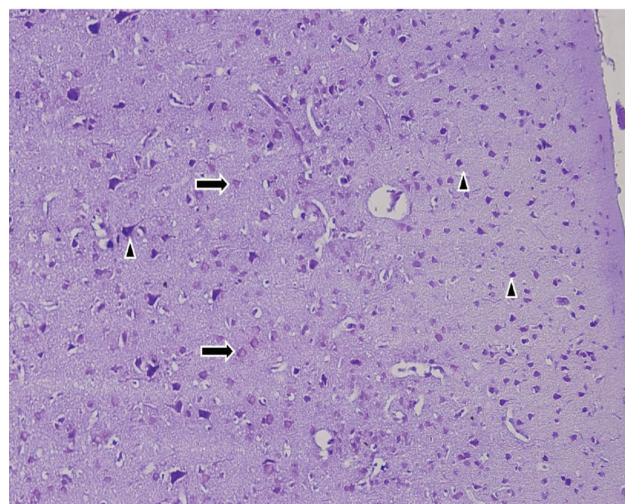


Figure 4. (Group-4) Cerebral cortex microphotographs (cresyl violet X 200). Normal pyramidal neurons (↑), degenerative pyramidal neurons (Δ)

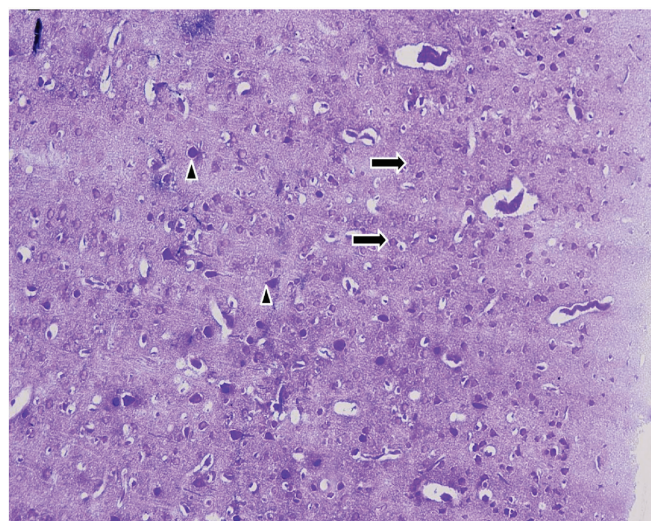


Figure 5. (Group-5) Cerebral cortex microphotographs (cresyl violet X 200). Normal pyramidal neurons (↑), degenerative pyramidal neurons (Δ)

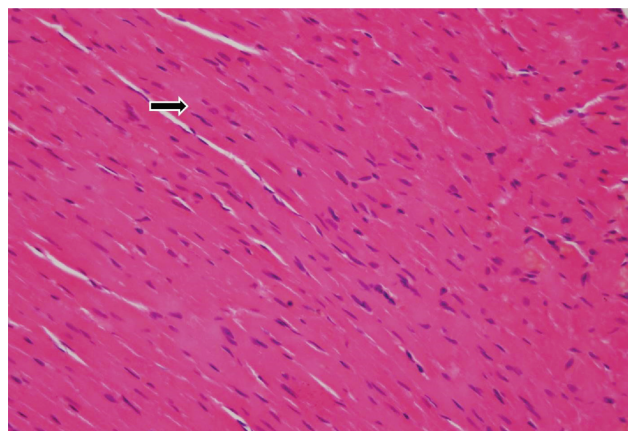


Figure 6. (Group-1) Myocardial tissue microphotographs (H&E X 200). Myocardial cells (↑)

congestion and edema between muscle fibers were observed (Figure 8). Occasional degeneration in muscle fibers, and diffuse vascular congestion and edema were observed in group 4

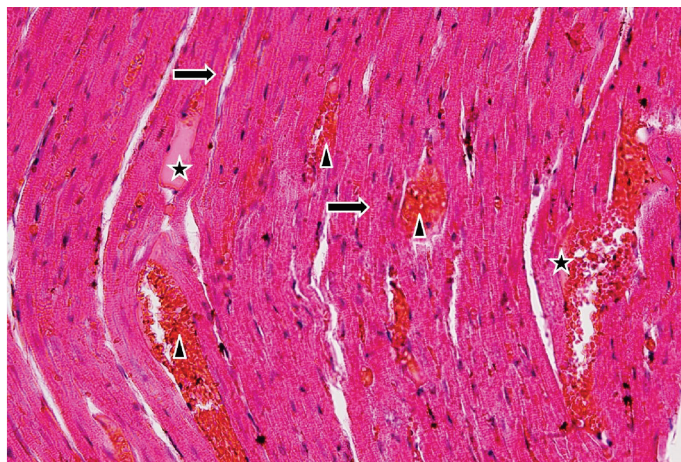


Figure 7. (Group-2) Myocardial tissue microphotographs (H&E X 200). Myocardial cells (↑), vascular congestion (Δ), edema (star)

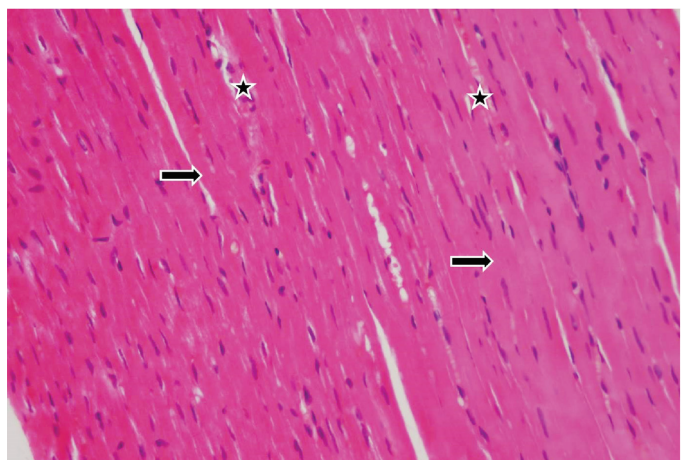


Figure 8. (Group-3) Myocardial tissue microphotographs (H&E X 200). Myocardial cells (↑), edema (star)

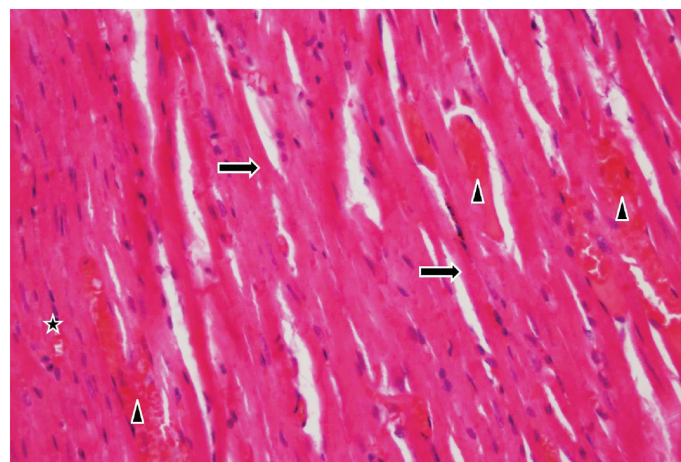


Figure 9. (Group-4) Myocardial tissue microphotographs (H&E X 200). Myocardial cells (↑), vascular congestion (Δ), edema (star)

(Figure 9). Cardiac muscle cells in group 5 exhibited a close to normal morphology, although occasional edema was observed between muscle fibers (Figure 10). Evaluation of hepatic tissues revealed a normal architecture in group 1. Normal morphology

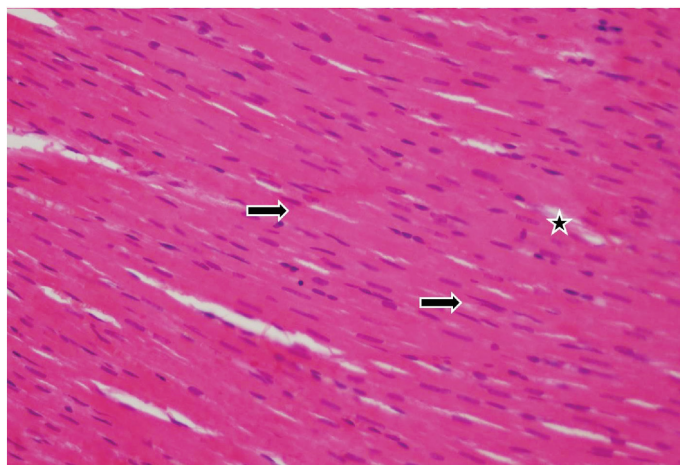


Figure 10. (Group-5) Myocardial tissue microphotographs (H&E X 200). Myocardial cells (↑), edema (star)

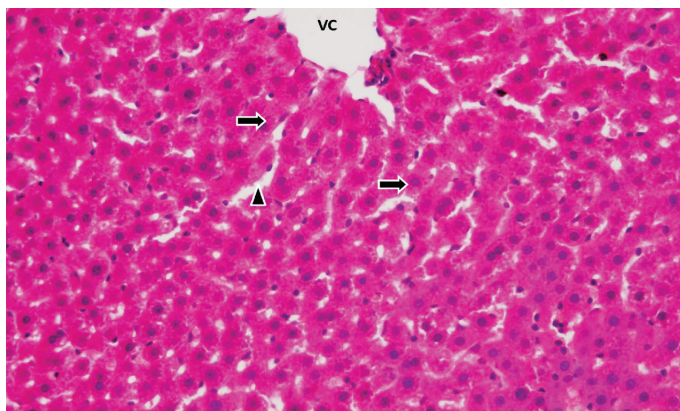


Figure 11. (Group-1) Liver tissue microphotographs (H&E X 200). Hepatocytes (↑), sinusoidal dilatation (Δ)

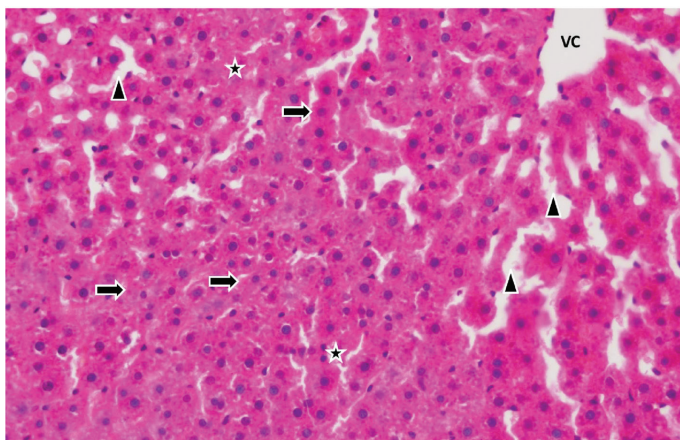


Figure 12. (Group-2) Liver tissue microphotographs (H&E X 200). Hepatocytes (↑), sinusoidal dilatation (Δ), hepatocytes with pyknotic nuclei (star)

was observed in hepatocytes and sinusoidal spaces (Figure 11). Diffuse hepatocytes with pyknotic nuclei and cell margin losses were observed in group 2. Diffuse expansion of sinusoidal spaces were also determined (Figure 12). Occasional hepatocytes with pyknotic nuclei and cell margin losses were observed in group 3. Occasional sinusoidal dilatation was observed in areas close to the vena centralis (Figure 13). Diffuse sinusoidal dilatation and degenerative hepatocytes with pyknotic nuclei were observed in group 4 (Figure 14). In group 5, diffuse hepatocytes with normal morphology were observed in addition to occasional hepatocytes with pyknotic nuclei (Figure 15).

Discussion

This study investigated the protective effects of EP and DMSO against injury caused in important target organs including liver, brain and heart by potentially life-threatening CO intoxication, which is frequently encountered worldwide. Our findings indicated that EP had no protective effect against histopathological damage occurring in brain, cardiac, and hepatic tissues. However, our findings also suggested that DMSO might exhibit protective effects against damage occurring in brain and cardiac tissue, but not against hepatic injury. However, this needs to be confirmed by further studies.

CO intoxication is a frequent health problem in worldwide. Approximately 40.000 people present to emergency departments in the USA every year due to CO intoxication, and 5000-6000 individuals die due to exposure to the gas. CO is held responsible for more than half of all fatal intoxications worldwide (24). The most common sources of CO are engine exhaust gases, gas-powered engines, smoke caused by fires, and paints containing methylene chloride (1).

The pathophysiology of CO intoxication is complex. In addition to the cellular damage caused by CO itself, the hypoxia and ischemia produced by the CO-hemoglobin complex also play an important role in the process (25). Endogenously produced CO results in 1% COHb. Low endogenous CO levels are physiological, but excessive exogenous CO intake causes the principal problem (26). The main target of CO is hemoglobin (Hb). The affinity of Hb for CO is 210 times greater than that for oxygen. CO binding to Hemoglobin (Hb) replaces the oxygen in Hb, giving rise to COHb. Since the oxygen binding regions on the COHb molecule have a high affinity for oxygen, the oxygen binded to these is not presented to tissues. This increased affinity is known as the Haldane effect. As a result of the Haldane effect, a left shift occurs on the oxyhemoglobin curve, and it becomes difficult for oxygen to separate from Hb at the tissue level (27). Tissue hypoxia develops since oxygen cannot be transported to tissues. Following recent studies, it is now thought that different mechanisms are also involved

in CO toxicity. One such is the damage occurring in the central nervous system with reoxygenation following hypoxia caused by CO. Measuring COHb levels is today considered an important

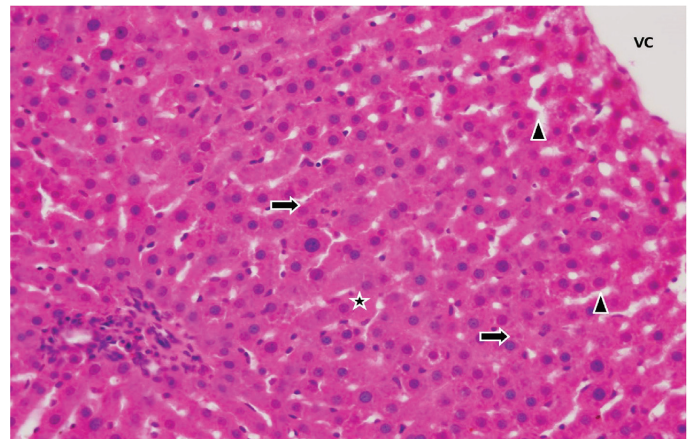


Figure 13. (Group-3) Liver tissue microphotographs (H&E X 200). Hepatocytes (↑), sinusoidal dilatation (Δ), hepatocytes with pyknotic nuclei (star)

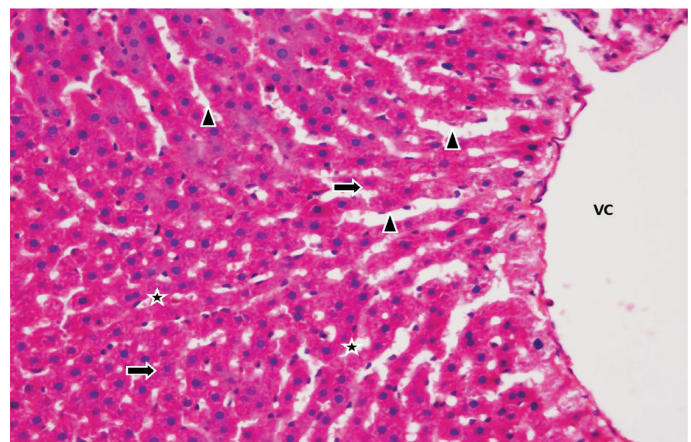


Figure 14. (Group-4) Liver tissue microphotographs (H&E X 200). Hepatocytes (↑), sinusoidal dilatation (Δ), hepatocytes with pyknotic nuclei (star)

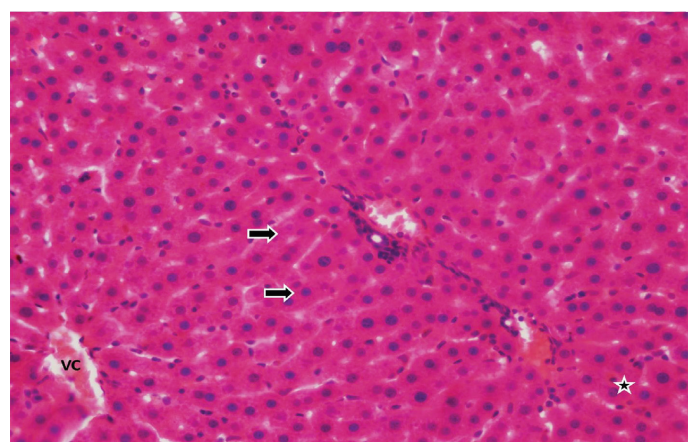


Figure 15. (Group-5) Liver tissue microphotographs (H&E X 200). Hepatocytes (↑), hepatocytes with pyknotic nuclei (star)

diagnostic technique. However, there is only a weak correlation between blood or tissue COHb levels and organ damage. Generally, COHb levels exceeding 60% result in mortality, while lower levels can result in clinical findings ranging from mild symptoms to coma, and it is not always possible to determine this by means of COHb levels (28). The therapeutic options in treatment planning for patients with CO intoxication are not numerous. Reactive oxygen product synthesis increases in CO intoxication secondary to tissue hypoxia. There is therefore a growing number of studies investigating the role and importance of antioxidant therapies as an alternative to oxygen therapy. In their experimental study, Wang et al. (29) showed that hydrogen-rich saline solution reduced lipid peroxidation in cerebral tissue, and might therefore have a positive effect on neurological sequelae. Another study determined that N-acetylcysteine (NAC) and melatonin reduced CO-induced brain and lung damage (30).

DMSO is an anti-inflammatory drug with a free oxygen radical (FOR) binding capacity (31). In clinical terms, it is used in the symptomatic treatment of interstitial cystitis (32). In experimental sepsis models, the inhibitory effects of DMSO on nuclear factor kappa receptors have been shown to regulate cellular gene expression in inflammation. It has been shown to exhibit a protective effect at the histological level with its powerful antioxidant activity in corrosive burns in experimental rat models (33,34).

EP is a promising antioxidant and anti-inflammatory agent with known neuroprotective efficacy. Pyruvate, the end product of glycolysis and an initial substrate of the tricarboxylic acid cycle, is the anionic form of 2-oxo-propionic acid. It has been reported to be capable of scavenging FORs and hydroxyl radicals. EP is the ethyl ester of pyruvate and has been reported to be a more powerful antioxidant than pyruvate (35).

To our knowledge, this is the first study investigating the protective effects of DMSO and EP on CO intoxication. DMSO is generally used in *in vivo* experiments as a solvent to dissolve chemicals. Because of this feature, it was used in the control group of many studies. In an experimental traumatic brain injury study, Di Giorgio et al. (36) wanted to evaluate the neuroprotective effect of curcumin and compare its effect with a known antioxidant α -tocopherol. DMSO was used as a solvent for curcumin in the treatment groups in this study. In the study there was both DMSO group and normal saline group for control. Surprisingly, Di Giorgio et al. (36) found that α -tocopherol, DMSO and all curcumin groups reduced the number of degenerating neurons compared to normal saline group and there was no significant difference between α -tocopherol, DMSO and curcumin groups. They concluded that, DMSO was an effective neuroprotective agent in experimental traumatic brain injury (36). Similarly, in

our study, we determined that DMSO had a therapeutic effect on ischemic injury.

There are several studies showing antioxidant and anti-inflammatory effects of EP. Suha et al. (37) investigated the protective effects of NAC and EP in an experimental electric burn model and found that EP had protective effects on heart and brain (37). But in our study, we observed EP alone did not have a therapeutic effect on CO intoxication. This may be due to the dose of EP administered. Administration of EP at different doses may have a protective effect. Further studies are needed to demonstrate this.

Our findings revealed a significant difference in all parameters between group 1 (SHAM) and group 4 (EP) ($p=0.004$), while no statistically significant difference was observed between group 2 (CO) and group 4. This suggested that EP alone had no therapeutic effect against histopathological injury. DMSO exhibited a greater capacity to reduce neuronal alteration grades, the degenerative neuron rates, and cardiac total injury score in group 3, the DMSO group, compared to group 2 ($p=0.005$, $p=0.002$, and $p=0.004$, respectively). Comparison between group 5 and group 2 revealed that EP + DMSO reduced the degenerative neuron rate, total cardiac injury score, and total hepatic injury score ($p=0.002$, $p=0.001$, and $p=0.004$, respectively). However, this treatment had no ameliorating effect on neuronal alteration grade. No significant difference was determined between group 3 and group 5 in terms of treatment results. In other words, DMSO administered alone was as effective as DMSO + EP. Comparison between group 1 and group 5 revealed no difference in terms of injury occurring in cardiac tissue, suggesting that the protective effect of EP + DMSO was more prominent against damage occurring in the heart.

Study Limitations

Due to ethical committee requirements, this study was performed with a limited number of rats. It therefore needs to be supported by further studies involving larger sample numbers. Although this was a controlled study, it may not have mimicked typical cases of CO intoxication seen in clinical practice. In addition, this study examined only histopathological damage resulting from high-dose CO exposure over 60 min and the therapeutic effect of antioxidants against that injury. Higher-dose intoxication concentrations were not investigated. The therapeutic agents EP and DMSO were administered at doses of 50 mg/kg, and 6 mg/kg, respectively, and the effectiveness of other doses could not be measured. In addition, although all animals inhaled CO in the same apparatus and for the same time, the same blood COHb level may not have been obtained in all rats.

Conclusion

Our findings showed that EP alone exhibited no protective effect on the organ injuries investigated, while DMSO exhibited a reducing effect on neuronal alteration grade, degenerative neuron rates, and total cardiac damage score. The ameliorating effect of combined treatment was more prominent in cardiac and hepatic injury.

Ethics

Ethics Committee Approval: Karadeniz Technical University Animal Care and Ethics Committee (approval no: 2016/23).

Informed Consent: Informed consent was not obtained due to the fact that the study is animal testing.

Peer-review: Externally peer-reviewed.

Authorship Contributions

Surgical and Medical Practices: S.P., M.İ., Ö.T., Concept: S.P., M.İ., Ö.T., Design: S.P., M.İ., Ö.T., Data Collection or Processing: M.Ç., M.Y., Analysis or Interpretation: M.Ç., M.Y., E.Y., Literature Search: S.P., M.İ., A.Ş., Y.K., Writing: S.P., M.İ., Ö.T.

Conflict of Interest: No conflict of interest was declared by the authors.

Financial Disclosure: The authors declared that this study received no financial support.

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